

Dramatic 5'-Residue Effect on Conformer Distribution of Short Oligonucleotide Retro Models of the Cisplatin–DNA Cross-Link: Implications for the Lippard and Cross-Link Distorted Base Pair Steps Present in Cisplatin–DNA Duplex Adducts

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An unusually distorted base pair (bp) step, which we refer to as the Lippard bp step, was discovered recently¹ next to the distorted G*G* step containing the adjacent intrastrand cross-link, the main DNA lesion formed by the anticancer drug cisplatin (G* = N7-platinated dG). The distortions arise mainly from a large movement of the 5'-G*•C bp relative to the X•X' and 3'-G*•C bps (Chart 1). The Lippard bp step was identified by solid-state X-ray methods in a DNA 16-mer duplex bound to an HMG protein.¹ By assessing in-depth NMR data, we concluded that the distorted step exists in all G*G* DNA duplexes characterized well by NMR methods.² The Lippard bp step is thus intrinsic to intrastrand lesions formed by this very widely used therapeutic drug.² Within the accuracy of the methods,² the distortions appear to involve the DNA, not the Pt geometry,³ and to be similar in solution and solid protein-bound states. Such good correlations between solution and solid-state results have been elusive because the dynamic nature and conformational diversity of DNA increase on cross-link formation, making structural assessments even more difficult.^{2,4}

Now that the distorted structure is well defined, we can compare features of the duplex with those of single-strand (ss) G*G* adducts to define the forces stabilizing the distortions. We chose to examine ss G*G* adducts with T residues because GGT is part of the repetitive sequence in ss telomeres, a potential cisplatin ss target.^{5,6} In solution, G*G* favors an HH1 form [head-to-head (HH) bases, *anti* G* residues, and the B-DNA phosphodiester backbone propagation direction]; this form is highly left-handed (L) canted in XG*G* ss^{7–9} and right-handed (R) canted in duplex adducts (Chart 2).⁴ This long-recognized difference in canting¹⁰ is poorly understood. Recent duplex studies^{1,2} provide the new information that the degree of R canting is low, and ammine H-bonds are either absent or very weak. The X residue has an N pucker,^{1,2,4} which may either cause the distortion exemplified by the Lippard bp step or be a consequence of the essentially normal X•X' WC H-bonding.^{1,2} In contrast, in L ss X-ray structures the ammine *cis* to the canted 5'-G* models always has an H-bond to the oligo,^{11,12} and the X residue has an S sugar.¹¹

The S pucker and the L canting in ss models^{7–9,11} may be favored by H-bonding with the carrier ligand; alternatively, the N pucker and R canting in the duplex may arise because such H-bonding is absent. Thus, it is of some interest to determine if the X-residue pucker and L canting are related to flanking residue H-bonding by examining Me₂ppzPt retro models because the Me₂ppz (*N,N'*-dimethylpiperazine) ligand cannot form NH H-bonds and does not

Chart 1. Distorted X•X', 5'-G*•C, and 3'-G*•C bp Region

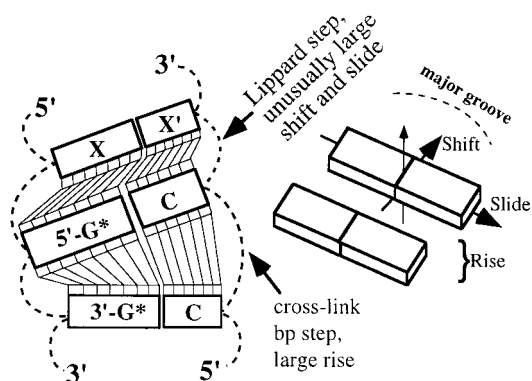
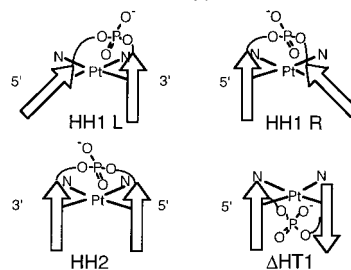


Chart 2. Known Forms of d(G*pG*) Retro Models Showing Both L-Canted (HH1 L) and R-canted (HH1 R) Forms (G base shown as an arrow with the H8 atom at tip)



influence conformer structure and distribution.¹³ Retro models, which have bulky carrier ligands, exhibit an $\sim 10^9$ reduction of the rotation rate of the guanine base relative to cisplatin,¹³ allowing us both to define structure and to identify new conformers of dinucleotide d(G*pG*) adducts in solution by NMR methods.^{4,14} (Phosphodiester linkage denoted by "p" only for dinucleotides.) The NMR data (Figure 1, Table 1, Supporting Information "SI") establish that Me₂ppzPt(d(TG*G*T)) exists at equilibrium as $\sim 100\%$ HH1 form. This is the first reported case in which an ss G*G* adduct has been shown to favor one conformer. Usually the HH1 form is in equilibrium with a second appreciable form, a ΔHT1 or an HH2 form (Chart 2).¹⁴ For Me₂ppzPt adducts at equilibrium, d(G*pG*)¹³ and d(G*G*T) uniquely have *three* significant forms, but, in contrast, d(TG*G*) has only HH1 (Table 1). Thus, a 3'-T has virtually no effect, but the 5'-T has a dramatic effect. *Carrier-ligand H-bonding cannot cause the high preference of the HH1 form because Me₂ppz cannot form H-bonds.* For all new adducts,

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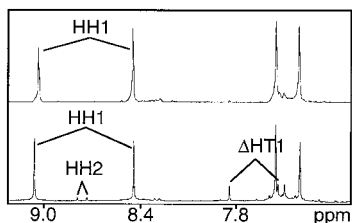


Figure 1. G H8/T H6 NMR signals for $\text{Me}_2\text{ppzPt}(\text{d}(\text{TG}^*\text{G}^*\text{T}))$, pH ~ 4 at ~ 14 days (bottom). Only the HH1 signals were present after ~ 60 days (top). Lines point to G H8 signals for each form.

Table 1. Distribution and NMR Data for Me_2ppzPt Adducts^a

Me_2ppzPt adduct	HH1 (%)	G*	H8 shifts (ppm) ^b	$J_{\text{H1}'-\text{H2}''}$ (Hz) ^b	$J_{\text{H1}'-\text{H2}''}$ (Hz) ^b
$\text{d}(\text{G}^*\text{pG}^*)^c$	50	5'	8.51	0	7.6
		3'	8.93	10.0	4.9
$\text{d}(\text{G}^*\text{G}^*\text{T})^c$	50	5'	8.48	0	8.3
		3'	8.93	9.2	4.8
$\text{d}(\text{TG}^*\text{G}^*\text{T})^d$	>97	5'	8.45	0	7.3
		3'	9.14	10.4	5.3
$\text{d}(\text{TG}^*\text{G}^*)^d$	>97	5'	8.47	0	7.5
		3'	9.10	9.8	5.0
$\text{d}(\text{pG}^*\text{pG}^*)^d$	>95	5'	8.59	0	7.2
		3'	9.02	9.1	4.7

^a D_2O at 5 °C, pH ~ 4 . ^b HH1. ^c $\sim 20\%$ HH2 and 30% ΔHT1 found after ~ 14 and ~ 60 days. ^d After ~ 60 days (or more); distributions at ~ 14 days were $\sim 80\%$ HH1, $\sim 4\text{--}8\%$ HH2, and $\sim 9\text{--}16\%$ ΔHT1 .

unambiguous NMR signals indicated formation of HH1, HH2, and ΔHT1 conformers as kinetic and sometimes equilibrium products (SI). This is the first reported characterization of multiple conformers in ss G^*G^* adducts longer than $\text{d}(\text{G}^*\text{pG}^*)$.

From downfield 3'- G^* H8 shifts of *cis*-Pt(NH₃)₂ ss adducts^{4,7–10} and X-ray structures [$\text{d}(\text{CG}^*\text{G}^*)$, three independent, very L-canted molecules with ammine 5'- G^* O6 H-bonding;¹¹ $\text{d}(\text{pG}^*\text{pG}^*)$, two molecules L with the 5'-p H-bonded¹²], L canting decreases in the order: 5'-residue $>$ 5'-p $>$ no substituent. This order is reflected in Me_2ppzPt adducts by very downfield 3'- G^* H8 NMR shifts (Table 1), clearly indicating that HH1 L canting decreases in the series: $\text{d}(\text{TG}^*\text{G}^*\text{T}) \approx \text{d}(\text{TG}^*\text{G}^*) > \text{d}(\text{pG}^*\text{pG}^*) > \text{d}(\text{G}^*\text{pG}^*) \approx \text{d}(\text{G}^*\text{G}^*\text{T})$. The degree of L canting clearly depends on 5'-X steric effects, not on ammine H-bonding.

From coupling data (SI), 5'-T of $\text{Me}_2\text{ppzPt}(\text{d}(\text{TG}^*\text{G}^*\text{T}))$ has an S pucker. Thus, the S pucker of the 5'-X residue is not related to ammine H-bonding. Indeed, by assessing models of hybrid structures constructed from pieces of known structures (SI), we find that L models with known C3'-O3' torsion angles (ϵ) have severe 5'-X 5'- G^* clashes unless the 5'-X has an S sugar pucker and $\epsilon = \sim -146^\circ$, the X-ray value for the highly L-canted *cis*-Pt(NH₃)₂- $\text{d}(\text{CG}^*\text{G}^*)$.¹¹ The latter has close 5'-X-5'- G^* contacts, consistent with the very upfield 5'-T H2' signals of Me_2ppzPt adducts (SI). A striking feature for all adducts is that the shifts and couplings of the G^*G^* sugar signals of the HH1 conformer are nearly identical when G^* lacks a flanking T (SI). Thus, the 5'-T residue, although positioned close to the G^*G^* lesion, causes no detectable changes in backbone geometry, consistent with the X-ray findings.^{11,12}

Hybrid models (SI) provide additional insight. In ss models, the severe clashes involving an L-canted 5'- G^* and an N-puckered 5'-X residue disappear for R canting. The 5'-X N pucker and the

backbone torsion angles in the Lippard and G^*G^* bp steps allow the X-X' bases to occupy positions favorable for stacking with the 5' flanking region. In this position, the 5'-X base does not clash with the 5'- G^* only when G^*G^* has R canting. (When the 5'-X in duplex hybrid models has the $\sim 180^\circ$ ϵ value of the distorted duplex,² R-canted 5'- G^* steric clashes with X are minimal even if the 5'-X has an S pucker, but the X and X' amino groups clash and no X-X' WC H-bonding is possible, SI.) The equal numbers of L- and R-canted molecules in the solid state for *cis*-Pt(NH₃)₂- $\text{d}(\text{pG}^*\text{pG}^*)$ ¹² and NMR data for $\text{d}(\text{G}^*\text{pG}^*)$ retro models¹⁴ provide evidence that L-to-R canting changes require little energy. The R molecules¹² simulate several aspects of the Lippard and G^*G^* bp steps^{1,2} (5'-p of 5'- G^* position and lack of NH₃ H-bonding, G^*G^* backbone structure, normal Pt geometry).

In conclusion, the highly L nature of ss adducts with a 5'-X residue is chiefly a consequence of the bulk of the 5' residue; H-bonding interactions are inconsequential. Any 5' substituent, including the 5'-p group, favors the HH1 form. However, the 5'-p group induces less L canting than a 5'-X residue. Most importantly, the 5' residue maintains an S pucker in ss adducts even in the absence of H-bonding, suggesting strongly that the N pucker of this residue in the Lippard bp step of duplexes is related to stacking and WC H-bonding, not to the insignificance of ammine H-bonding. Hybrid models built with the *cis*-Pt(NH₃)₂- $\text{d}(\text{pG}^*\text{pG}^*)$ X-ray structure¹² reveal that R canting effectively eliminates clashes with the 5'-X of an X-X' bp. Our analysis rationalizes key solid-state and solution data in a satisfying fashion.

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Supporting Information Available: Tables and figures of NMR data and of superimposed structures derived from X-ray and NMR data (PDF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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